

TOPICAL REVIEW

The microbiota–gut–brain axis in gastrointestinal disorders: stressed bugs, stressed brain or both?

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Abstract The gut–brain axis is the bidirectional communication between the gut and the brain, which occurs through multiple pathways that include hormonal, neural and immune mediators. The signals along this axis can originate in the gut, the brain or both, with the objective of maintaining normal gut function and appropriate behaviour. In recent years, the study of gut microbiota has become one of the most important areas in biomedical research. Attention has focused on the role of gut microbiota in determining normal gut physiology and immunity and, more recently, on its role as modulator of host behaviour ('microbiota–gut–brain axis'). We therefore review the literature on the role of gut microbiota in gut homeostasis and link it with mechanisms that could influence behaviour. We discuss the association of dysbiosis with disease, with particular focus on functional bowel disorders and their relationship to psychological stress. This is of particular interest because exposure to stressors has long been known to increase susceptibility to and severity of gastrointestinal diseases.

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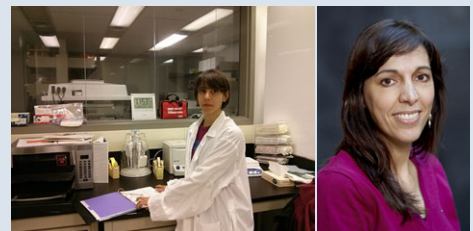
Introduction

The central nervous system and the gastrointestinal (GI) tract are in constant bidirectional communication through neural pathways, such as the vagus nerve, and by humoral and cellular mediators that include the immune system and the hypothalamic–pituitary–adrenal (HPA) axis.

The gut is colonized with a complex community of bacteria (microbiota), which helps to shape the immune system, metabolic function and behaviour in health and disease throughout life. The microbiota is a relatively new player in the gut–brain axis, fulfilling key roles in its communication (Bailey & Coe, 1999; Bercik *et al.* 2011a; Heijtz *et al.* 2011; Neufeld *et al.*

2011; Matsumoto *et al.* 2013), which has led to the term 'microbiota–gut–brain axis' (Rhee *et al.* 2009; Collins *et al.* 2012). Alterations in gut microbiota (dysbiosis) can arise as a consequence of gastrointestinal disease or of its treatment. All major chronic disorders of the gut, namely inflammatory bowel disease, irritable bowel syndrome and coeliac disease, are associated with dysbiosis (Nadal *et al.* 2007; Collado *et al.* 2009; De Palma *et al.* 2010). Although an overall decrease in diversity and richness of the microbiota seems to be a common finding across studies, no specific dysbiotic signature has emerged between studies. This may be due, in part, to differences in sampling (small intestinal, colonic, faecal), as well as analytical techniques employed (culture, Denaturing

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Gradient Gel Electrophoresis, DGGE, Illumina, 454 sequencing, Matrix-assisted laser desorption ionization time-of-flight mass spectrometer, MALDI-TOF) (Lagier *et al.* 2012). However, there is now increasing evidence that dysbiosis modulates peripheral and central nervous system function, leading to alterations in brain signalling and behaviour (Bercik *et al.* 2011a; Collins *et al.* 2013; Mulle *et al.* 2013). This observation is important in view of the fact that stress and depression, common co-morbidities in GI disorders, in turn influence the natural course of these illnesses (Collins, 2001; Wu, 2012).

The microbiota–gut–brain axis has been the subject of numerous reviews in recent years (Rhee *et al.* 2009; Mayer, 2011, 2014; Bercik *et al.* 2012; Collins *et al.* 2012, 2013). The influence of psychosocial and environmental stressors on the pathogenesis of gastrointestinal diseases has long been recognized. Recently, the mechanisms through which stress may affect various physiological functions of the GI tract have been reviewed (Konturek *et al.* 2011). We will discuss the recent progress on specific mechanisms of interaction between gut microbiota and brain, with focus on the effect of psychological stress.

Gut microbiota and its host: a mutualistic relationship

A unique combination of different populations of organisms inhabits our gut, mainly bacteria but also archaea, viruses and protozoa, roughly approximating 10^{14} cells, outnumbering the human cells in our bodies by a factor of 10 (Sekirov *et al.* 2010). While bacterial profiling and its understanding has become easier during the last decade, the analysis of the mycobiome and the virome is still in its infancy (Minot *et al.* 2011, 2013; Cui *et al.* 2013).

The human intestinal tract is essentially sterile at birth, when it is immediately colonized. The gut microbiota evolves during early life until a unique, subject-specific (fingerprint) adult-like community arises, which is relatively stable throughout life (Rajilić-Stojanović *et al.* 2013). Out of the more than 50 phyla described in the literature, only few are found in the human GI tract, dominated by two phyla in particular (Firmicutes, Bacteroidetes), together with members of Actinobacteria, Verrucomicrobia, Proteobacteria, Fusobacteria and Cyanobacteria phyla (Sommer & Bäckhed, 2013). These autochthonous phyla colonize the GI tract and are present in a majority of individuals. The concept of ‘enterotypes’ has recently been proposed and, according to this, humans can be subdivided into *Bacteroides*, *Prevotella* or *Ruminococcus* types (Zoetendal *et al.* 2008; Arumugam *et al.* 2011). However, this categorization has recently become a matter of debate, and the term ‘enterogradient’ has been proposed instead, to describe bacterial communities with prevalence of

Bacteroides or *Prevotella* (Jeffery *et al.* 2012). Microbes in the human gut undergo selective pressure from the host as well as from microbial competitors, and once the ecosystems reaches homeostasis, some species will occur in high and many in low abundance (Bäckhed, 2011; Nicholson *et al.* 2012).

Even though the gut microbiota differs greatly between subjects in membership and community structure, it still appears on the whole to be functionally equivalent and necessary for the proper development of the host. Mammals have co-evolved to exist with their gut microbiota largely in a mutualistic relationship; these organisms participate in the conversion of non-digestible carbohydrates (dietary fibre) to short-chain fatty acids, participate in bile acid metabolism, provide a barrier against pathogenic bacteria, and modulate the innate and adaptive immune systems (Nicholson *et al.* 2012). In turn, the host provides a unique, nutrient-rich niche at constant temperature (Sommer & Bäckhed, 2013).

Studies using germ-free animals have highlighted the importance of the gut microbiota in the maintenance of homeostasis. Germ-free animals have physiological and metabolic abnormalities compared with conventional animals, as well as an imbalanced immune system (Slack *et al.* 2009; Hapfelmeier *et al.* 2010; Geuking *et al.* 2011; Kunii *et al.* 2011; Hansen *et al.* 2012; Macpherson *et al.* 2012; Olszak *et al.* 2012). In addition, germ-free animals exhibit abnormal gastrointestinal motility (Abrams & Bishop, 1967; Gustafsson *et al.* 1970; Wostmann, 1981), increased expression of genes encoding transporters throughout the gut (Bäckhed, 2011) and altered perception of inflammatory pain (Amaral *et al.* 2008). Moreover, germ-free mice have an impaired capacity to harvest energy from the diet (Wostmann, 1981) and are protected against diet-induced obesity (Bäckhed *et al.* 2007; Rabot *et al.* 2010).

It is therefore not surprising that alterations in the composition of the normal gut microbiota (dysbiosis) are associated with a variety of GI disorders, such as inflammatory bowel diseases, irritable bowel syndrome and coeliac disease (Nadal *et al.* 2007; Collado *et al.* 2009; De Palma *et al.* 2010). Future work will have to determine whether a microbial signature for dysbiosis is associated with specific disease states. Nevertheless, sufficient data support the concept that changes in the microbiota may arise in adulthood as a consequence of disease, long-term dietary habits, antibiotics and medications. These changes may be short term or long term, depending on the duration of the trigger that induced them and the particular characteristics of the host. In contrast, factors that impact on the normal colonization process during early life, such as psychological stress, may exert long-term effects on the composition of the microbiota that will impact susceptibility to disease.

Microbiota–gut–brain axis

It is well known that the gut and the brain are in bidirectional communication. The concept of the gut–brain axis originated from the field of GI endocrinology and the discovery of hormonal regulation of digestion (Track, 1980). Since then, it has evolved to include the maintenance of homeostasis of several systems, including GI function, appetite and weight control (Collins & Bercik, 2009). Thus, it is only logical to consider and include the gut microbiota as an important modulator of this system and, consequently, the term ‘microbiota–gut–brain axis’ has emerged (Fig. 1; Bercik *et al.* 2009, 2011a).

The known beneficial effects of laxatives and oral antibiotics in patients with hepatic encephalopathy is perhaps one of the earliest pieces of evidence for a role of gut bacteria in brain function (Victor & Quigley, 2014). Antibiotics have also anecdotally been reported to induce acute psychosis that resolved after withdrawal of the drug (Sternbach & State, 1997; Mehdi, 2010). More recently, an abnormal composition of the microbiota has been associated with autism (Bolte, 1998; Finegold *et al.* 2010, 2012; Yap *et al.* 2010; Wang *et al.* 2011, 2012, 2013; Williams *et al.* 2011, 2012; De Angelis *et al.* 2013; Kang *et al.* 2013); treatment with antibiotics in patients with late-onset autism seems to improve their symptoms (Sandler *et al.* 2000; Finegold *et al.* 2012). *Bacteroides fragilis*, a Gram-negative anaerobic bacterium that inhabits the lower GI tract of most mammals (Ley

et al. 2008), has been shown to ameliorate anxiety-like behavior, sensorimotor, communicative and repetitive behavior, but not sociability and social preference in an animal model of autism. The underlying mechanisms may involve modulation of gut microbiota composition and serum metabolomic profile (Hsiao *et al.* 2013). An association between major depressive disorder and altered gut metabolism has also been proposed (Ledochowski *et al.* 1998a,b, 2000, 2001; Ochoa-Repáraz *et al.* 2011).

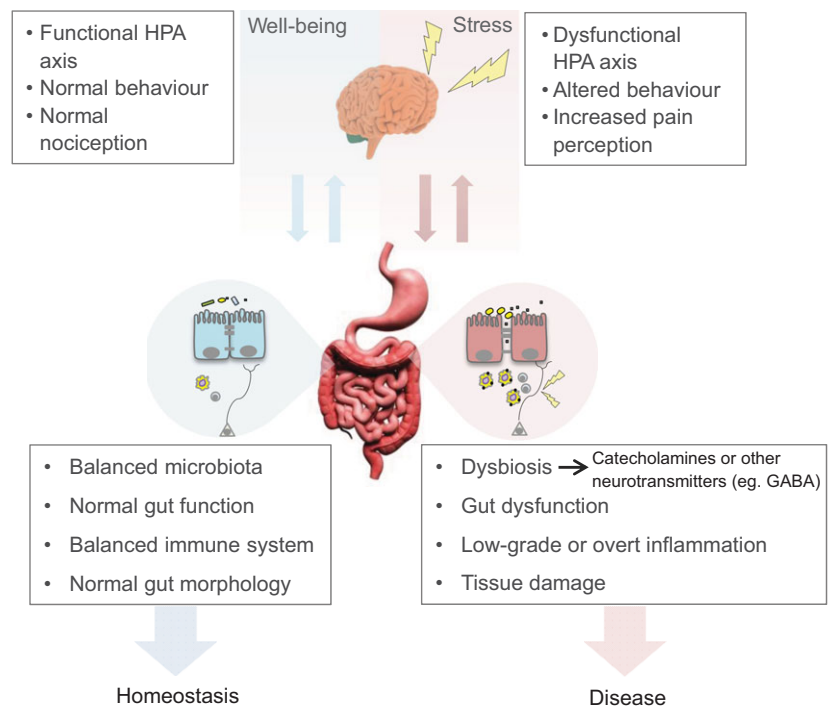
It is difficult to interpret whether this is a chicken or egg situation, whether brain and behavioural alterations precede gut dysfunction and dysbiosis, or whether gut dysfunction and dysbiosis precede brain and behavioural changes. It has been reported that chronic depression is associated with altered microbial profiles and colonic motility in mice (Park *et al.* 2013). However, it has been also reported that chronic gastrointestinal inflammation can induce anxiety-like behaviour and alter central nervous system biochemistry (Bercik *et al.* 2010, 2011a). Therefore, it is likely that both situations coexist in a self-perpetuating loop, and that the initial trigger can arise centrally or in the periphery. Additional research is needed to solve this intriguing concept, and an interaction between clinical and basic research using gnotobiotic technology will probably help to provide mechanistic insight.

Stress and the microbiota–gut–brain axis

Stress is defined as an organism’s total response to environmental demands or pressures. Several different

Figure 1. The microbiota–gut–brain axis comprises the bidirectional communication, through multiple pathways, between the gut and the brain

During stress, alterations at the level of the central nervous system can influence gut neuromotor and secretory function, immunity and microbiota composition. In turn, dysbiosis may contribute to perpetuate dysfunction and inflammation, further disrupting gut–brain communication. Some of these effects may be mediated by direct host–microbial interactions at the level of the intestinal epithelium, production of bacterial metabolites (catecholamines, GABA, etc). The sequence of events can occur in a top-to-bottom or bottom-to-top fashion, but once initiated can perpetuate and exacerbate maladaptive responses that promote a state of disease. We acknowledge dreamdesign and cooldesign (FreeDigitalPhotos.net) for the image of the gut and brain, respectively.



types of stressors can be distinguished, such as acute or chronic, some of which may occur only once, while others are repetitive and can be anticipated. However, stress can be unpredictable and uncontrollable, mild or severe, and occur in or out of context (Lucassen *et al.* 2014). Moreover, the perception of stress is variable between individuals, and so is the persistence of its consequences (Lucassen *et al.* 2014). Exposure to stressors has long been known to increase susceptibility to disease, including GI disorders. Stress contributes to many disabilities worldwide and, as such, represents a severe economic burden.

Chronic and acute stress models are widely employed in GI research, because stress has been identified as a risk factor or modulator of the expression of several GI disorders (Collins, 2001; Söderholm & Perdue, 2001; Konturek *et al.* 2011). Tannock and Savage demonstrated, 40 years ago, that environmental and dietary stress markedly altered the gut microbiota in mice, affecting factors that regulate the localization and population levels of micro-organisms along the GI tract (Tannock & Savage, 1974), possibly favouring the establishment of pathogenic bacterial species (Tannock & Smith, 1972; Tannock & Savage, 1974). More recently, Bailey *et al.* (2011) demonstrated that exposure to a social disruption stressor affects the gut microbiota and circulating levels of cytokines, particularly interleukin-6 and monocyte chemoattractant protein-1. In fact, social stress has been reported to increase the risk of inflammation-related diseases, promoting pro-inflammatory gene expression and monocyte differentiation (Powell *et al.* 2013). Thus, stressor-induced changes in the microbiota may enhance the ability of enteric pathogens (such as *Citrobacter rodentium*) to colonize the intestine (Bailey *et al.* 2010). Accordingly, it has been reported that acute and repeated stress affect levels of intestinal secretory IgA, impacting intestinal homeostasis and probably resulting in inflammation (Campos-Rodríguez *et al.* 2013). Altered levels of intestinal secretory IgA might cause shifts in commensals and possibly result in dysbiosis.

Psychological and physical stressors activate the HPA axis, resulting in the release of corticotrophin-releasing hormone, the principal regulator of the HPA axis, which is synthesized and secreted by hypophysiotrophic neurons localized in the medial parvocellular subdivision of the paraventricular nucleus (Smith & Vale, 2006). Corticotrophin-releasing hormone induces the release of adrenocorticotrophic hormone into the systemic circulation, which will, in turn, stimulate glucocorticoid synthesis in the adrenal cortex. Glucocorticoids, such as corticosterone or cortisol in humans, are the downstream effectors of the HPA axis, and their biological effects are usually adaptive (Smith & Vale, 2006). Together with glucocorticoids, catecholamines (noradrenaline and adrenaline) are also released into the circulatory system after psychological and physical stressors (Lyte *et al.* 2011),

and it is well known that glucocorticoids can potentiate some of the actions of catecholamines (Sapolsky *et al.* 2000).

The gastrointestinal tract has long been known to be sensitive to stress and stress mediators, including catecholamines, but the notion that stress, and stress mediators, can influence the composition and function of the gut microbiota is a relatively new concept (Lyte *et al.* 2011). In fact, stress can influence the outcome of bacterial infection, because enteric bacteria can respond to the release of stress-related neurochemical mediators by the host (Lyte *et al.* 2011). Moreover, it has been hypothesized recently that bacteria act essentially as neuro-active compound delivery vehicles, affecting host physiology through the provision of neurochemicals. Specifically, the presence of a stress-related neuroendocrine hormone family of catecholamines has been demonstrated in bacteria (Lyte, 2011).

Today's conceptual framework of the most common entities in gastroenterology, the functional gastrointestinal disorders, such as irritable bowel syndrome and functional dyspepsia, involves the interaction of psychological factors and altered gut physiology via the gut–brain axis, where brain and gut symptoms are reciprocally influencing each other's expression. Psychological, sexual and/or physical abuse in early life has been suggested to play an important role in the pathogenesis of functional gastrointestinal disorders (Heitkemper *et al.* 2011; Wu, 2012; van Tilburg *et al.* 2013). This is a time of particular vulnerability, when neurological plasticity as well as establishment of a relatively stable gut microbiota occurs.

Maternal separation in rodents has been widely used as a model of early life stress that induces long-lasting hyperactivity of the HPA axis (Ladd *et al.* 2000; Barreau *et al.* 2004b; Daniels *et al.* 2004; Lippmann *et al.* 2007; Aisa *et al.* 2008; Gareau *et al.* 2008; Oines *et al.* 2012), anxiety-like behaviour (Varghese *et al.* 2006; Lippmann *et al.* 2007; Desbonnet *et al.* 2010; O'Mahony *et al.* 2011; Abelaira *et al.* 2013; Li *et al.* 2013), visceral hypersensitivity (Eutamene *et al.* 2007; O'Mahony *et al.* 2011; Moloney *et al.* 2012; Felice *et al.* 2014) and altered cholinergic activity in the gut (Gareau *et al.* 2007b; O'Malley *et al.* 2010) accompanied by increased intestinal permeability (Söderholm *et al.* 2002; Barreau *et al.* 2004a; García-Ródenas *et al.* 2006; Eutamene *et al.* 2007; Gareau *et al.* 2007b; Oines *et al.* 2012).

Maternally separated rats show also increased neuronal activation in response to a physical stressor, such as colorectal distension (Felice *et al.* 2014), probably due to central sensitization to noxious visceral stimuli (Chung *et al.* 2007), similar to what has been reported for irritable bowel syndrome patients (Tillisch & Labus, 2011; Tillisch *et al.* 2011; Larsson *et al.* 2012). Indeed, this model results in a dysfunctional gut–brain axis, mimicking many of the features found in irritable bowel syndrome

patients; therefore, it has been widely employed to study the mechanisms behind the dysfunctional communication between the gut and the brain in irritable bowel syndrome (Barreau *et al.* 2007; Gareau *et al.* 2008; O'Mahony *et al.* 2009, 2011). Similar to irritable bowel syndrome (Ringel & Maharshak, 2013), in animal models these alterations at physiological and behavioural levels are often accompanied by altered gut colonization (García-Ródenas *et al.* 2006; O'Mahony *et al.* 2009; Barouei *et al.* 2012), and the use of probiotics appears to improve the detrimental effects of stress (García-Ródenas *et al.* 2006; Eutamene & Bueno, 2007; Eutamene *et al.* 2007; Gareau *et al.* 2007a; Desbonnet *et al.* 2010; Distrutti *et al.* 2013).

Our preliminary data show that gut microbiota is essential for the expression of anxiety-like behaviour and behavioural despair in mice, because maternally separated germ-free mice do not show different behaviour when compared with control germ-free mice (De Palma *et al.* 2012). However, we found that germ-free maternally separated mice have increased levels of basal serum corticosterone and altered cholinergic nerve function (De Palma *et al.* 2012), similar to previous studies in conventional specific pathogen-free animals (Gareau *et al.* 2006, 2007a,b; O'Malley *et al.* 2011), indicating that these alterations occur independently of the presence of gut microbiota.

Acetylcholine is the main excitatory neurotransmitter in the mammalian enteric nervous system and plays an important role in the control of gut motility (Olsson & Holmgren, 2011). Park *et al.* (2013) demonstrated that central administration of corticotrophin-releasing hormone induces changes in colonic motility in mice, accompanied by altered behaviour in the open field test. Thus, change in the HPA axis may contribute to the development of diverse pathologies; in this case, it altered autonomic control of gut motility (Park *et al.* 2013). We obtained similar results in germ-free mice subjected to maternal separation, demonstrating that alterations at the level of HPA axis activity disrupt colonic homeostasis and, in turn, alter the gut environment, in a microbiota-independent fashion.

Maternal separation also induces changes in the morphology of the colon of conventional specific pathogen-free maternally separated rats, with an increase in the numbers of goblet cells in the crypts of the proximal colon and a subsequent increase in secretion of mucus, with a thinner mucosal layer (O'Malley *et al.* 2010). It is therefore plausible that changes to the physiology (Söderholm *et al.* 2002; Gareau *et al.* 2007b; O'Malley *et al.* 2010; De Palma *et al.* 2012) and morphology (O'Malley *et al.* 2010) of the gut of maternally separated animals explain the reported changes in gut microbiota composition of maternally separated animals *versus* control animals (O'Mahony *et al.* 2009).

Altogether, these findings suggest that stress, whether acute or chronic, modulates the gut environment to select a dysbiotic microbiota, which in turn can induce anxiety and depression; however, the exact pathways and mediators of this effect are yet to be elucidated. Commensal bacteria might modulate brain biochemistry and behaviour through the production of specific metabolites (Lyte, 2011; Barrett *et al.* 2012a,b; Hsiao *et al.* 2013). It has been shown previously that commensal bacteria can modulate behaviour through the vagus nerve (Bercik *et al.* 2011b; Bravo *et al.* 2011), affecting neurotransmitter metabolism (Asano *et al.* 2012), or through alternative pathways, yet to be defined (Bercik *et al.* 2010, 2011a).

It is plausible to postulate that in the future the manipulation of gut microbiota, through probiotics or symbiotics, might be a valuable adjuvant to traditional medicine in the treatment of irritable bowel syndrome patients with co-morbid anxiety or depression.

References

- Abelaira HM, Réus GZ & Quevedo J (2013). Animal models as tools to study the pathophysiology of depression. *Rev Bras Psiquiatr* **35**(Suppl 2), S112–S120.
- Abrams GD & Bishop JE (1967). Effect of the normal microbial flora on gastrointestinal motility. *Proc Soc Exp Biol Med* **126**, 301–304.
- Aisa B, Tordera R, Lasheras B, Del Río J & Ramírez MJ (2008). Effects of maternal separation on hypothalamic–pituitary–adrenal responses, cognition and vulnerability to stress in adult female rats. *Neuroscience* **154**, 1218–1226.
- Amaral FA, Sachs D, Costa VV, Fagundes CT, Cisalpino D, Cunha TM, Ferreira SH, Cunha FQ, Silva TA, Nicoli JR, Vieira LQ, Souza DG & Teixeira MM (2008). Commensal microbiota is fundamental for the development of inflammatory pain. *Proc Natl Acad Sci U S A* **105**, 2193–2197.
- Arumugam M, Raes J, Pelletier E, Le PD, Yamada T, Mende DR, Fernandes GR, Tap J, Bruls T, Batto JM, Bertalan M, Borruel N, Casellas F, Fernandez L, Gautier L, Hansen T, Hattori M, Hayashi T, Kleerebezem M, Kurokawa K, Leclerc M, Levenez F, Manichanh C, Nielsen HB, Nielsen T, Pons N, Poulain J, Qin J, Sicheritz-Ponten T, Tims S, Torrents D, Ugarte E, Zoetendal EG, Wang J, Guarner F, Pedersen O, de Vos WM, Brunak S, Doré J, Antolín M, Artiguenave F, Blottiere HM, Almeida M, Brechot C, Cara C, Chervaux C, Cultrone A, Delorme C, Denariuz G, Dervyn R, Foerstner KU, Friss C, van de Guchte M, Guedon E, Haimet F, Huber W, van Hylckama-Vlieg J, Jamet A, Juste C, Kaci G, Knol J, Lakhdari O, Layec S, Le RK, Maguin E, Mérieux A, Melo MR, M'rini C, Muller J, Oozeer R, Parkhill J, Renault P, Rescigno M, Sanchez N, Sunagawa S, Torrejon A, Turner K, Vandemeulebrouck G, Varela E, Winogradsky Y, Zeller G, Weissenbach J, Ehrlich SD & Bork P (2011). Enterotypes of the human gut microbiome. *Nature* **473**, 174–180.

- Asano Y, Hiramoto T, Nishino R, Aiba Y, Kimura T, Yoshihara K, Koga Y & Sudo N (2012). Critical role of gut microbiota in the production of biologically active, free catecholamines in the gut lumen of mice. *Am J Physiol Gastrointest Liver Physiol* **303**, G1288–G1295.
- Bäckhed F (2011). Programming of host metabolism by the gut microbiota. *Ann Nutr Metab* **58**(Suppl 2), 44–52.
- Bäckhed F, Manchester JK, Semenkovich CF & Gordon JI (2007). Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc Natl Acad Sci U S A* **104**, 979–984.
- Bailey MT & Coe CL (1999). Maternal separation disrupts the integrity of the intestinal microflora in infant rhesus monkeys. *Dev Psychobiol* **35**, 146–155.
- Bailey MT, Dowd SE, Galley JD, Hufnagle AR, Allen RG & Lyte M (2011). Exposure to a social stressor alters the structure of the intestinal microbiota: implications for stressor-induced immunomodulation. *Brain Behav Immun* **25**, 397–407.
- Bailey MT, Dowd SE, Parry NM, Galley JD, Schauer DB & Lyte M (2010). Stressor exposure disrupts commensal microbial populations in the intestines and leads to increased colonization by *Citrobacter rodentium*. *Infect Immun* **78**, 1509–1519.
- Barouei J, Moossavi M & Hodgson DM (2012). Effect of maternal probiotic intervention on HPA axis, immunity and gut microbiota in a rat model of irritable bowel syndrome. *PLoS One* **7**, e46051.
- Barreau F, Cartier C, Ferrier L, Fioramonti J & Bueno L (2004a). Nerve growth factor mediates alterations of colonic sensitivity and mucosal barrier induced by neonatal stress in rats. *Gastroenterology* **127**, 524–534.
- Barreau F, Ferrier L, Fioramonti J & Bueno L (2004b). Neonatal maternal deprivation triggers long term alterations in colonic epithelial barrier and mucosal immunity in rats. *Gut* **53**, 501–506.
- Barreau F, Ferrier L, Fioramonti J & Bueno L (2007). New insights in the etiology and pathophysiology of irritable bowel syndrome: contribution of neonatal stress models. *Pediatr Res* **62**, 240–245.
- Barrett E, Fitzgerald P, Dinan TG, Cryan JF, Ross RP, Quigley EM, Shanahan F, Kiely B, Fitzgerald GF, O'Toole PW & Stanton C (2012a). Bifidobacterium breve with α -linolenic acid and linoleic acid alters fatty acid metabolism in the maternal separation model of irritable bowel syndrome. *PLoS One* **7**, e48159.
- Barrett E, Ross RP, O'Toole PW, Fitzgerald GF & Stanton C (2012b). γ -Aminobutyric acid production by culturable bacteria from the human intestine. *J Appl Microbiol* **113**, 411–417.
- Bercik P, Collins SM & Verdu EF (2012). Microbes and the gut-brain axis. *Neurogastroenterol Motil* **24**, 405–413.
- Bercik P, Denou E, Collins J, Jackson W, Lu J, Jury J, Deng Y, Blennerhassett P, Macri J, McCoy KD, Verdu EF & Collins SM (2011a). The intestinal microbiota affect central levels of brain-derived neurotrophic factor and behavior in mice. *Gastroenterology* **141**, 599–609, 609e.1–3.
- Bercik P, Park AJ, Sinclair D, Khoshdel A, Lu J, Huang X, Deng Y, Blennerhassett PA, Fahnestock M, Moine D, Berger B, Huizinga JD, Kunze W, McLean PG, Bergonzelli GE, Collins SM & Verdu EF (2011b). The anxiolytic effect of *Bifidobacterium longum* NCC3001 involves vagal pathways for gut–brain communication. *Neurogastroenterol Motil* **23**, 1132–1139.
- Bercik P, Verdu EF, Foster JA, Lu J, Scharringa A, Kean I, Wang L, Blennerhassett P & Collins SM (2009). Role of gut-brain axis in persistent abnormal feeding behavior in mice following eradication of *Helicobacter pylori* infection. *Am J Physiol Regul Integr Comp Physiol* **296**, R587–R594.
- Bercik P, Verdu EF, Foster JA, Macri J, Potter M, Huang X, Malinowski P, Jackson W, Blennerhassett P, Neufeld KA, Lu J, Khan WI, Cortesey-Theulaz I, Cherbut C, Bergonzelli GE & Collins SM (2010). Chronic gastrointestinal inflammation induces anxiety-like behavior and alters central nervous system biochemistry in mice. *Gastroenterology* **139**, 2102–2112.
- Bolte ER (1998). Autism and *Clostridium tetani*. *Med Hypotheses* **51**, 133–144.
- Bravo JA, Dinan TG & Cryan JF (2011). Alterations in the central CRF system of two different rat models of comorbid depression and functional gastrointestinal disorders. *Int J Neuropsychopharmacol* **14**, 666–683.
- Campos-Rodríguez R, Godínez-Victoria M, Abarca-Rojano E, Pacheco-Yépez J, Reyna-Garfias H, Barbosa-Cabrera RE & Drago-Serrano ME (2013). Stress modulates intestinal secretory immunoglobulin A. *Front Integr Neurosci* **7**, 86.
- Chung EK, Zhang X, Li Z, Zhang H, Xu H & Bian Z (2007). Neonatal maternal separation enhances central sensitivity to noxious colorectal distention in rat. *Brain Res* **1153**, 68–77.
- Collado MC, Donat E, Ribes-Koninckx C, Calabuig M & Sanz Y (2009). Specific duodenal and faecal bacterial groups associated with paediatric coeliac disease. *J Clin Pathol* **62**, 264–269.
- Collins SM (2001). Stress and the gastrointestinal tract IV. Modulation of intestinal inflammation by stress: basic mechanisms and clinical relevance. *Am J Physiol Gastrointest Liver Physiol* **280**, G315–G318.
- Collins SM & Bercik P (2009). The relationship between intestinal microbiota and the central nervous system in normal gastrointestinal function and disease. *Gastroenterology* **136**, 2003–2014.
- Collins SM, Kassam Z & Bercik P (2013). The adoptive transfer of behavioral phenotype via the intestinal microbiota: experimental evidence and clinical implications. *Curr Opin Microbiol* **16**, 240–245.
- Collins SM, Surette M & Bercik P (2012). The interplay between the intestinal microbiota and the brain. *Nat Rev Microbiol* **10**, 735–742.
- Cui L, Morris A & Ghedin E (2013). The human mycobiome in health and disease. *Genome Med* **5**, 63.
- Daniels WM, Pietersen CY, Carstens ME & Stein DJ (2004). Maternal separation in rats leads to anxiety-like behavior and a blunted ACTH response and altered neurotransmitter levels in response to a subsequent stressor. *Metab Brain Dis* **19**, 3–14.

- De Angelis M, Piccolo M, Vannini L, Siragusa S, De Giacomo A, Serrazzanetti DI, Cristofori F, Guerzoni ME, Gobetti M & Francavilla R (2013). Fecal microbiota and metabolome of children with autism and pervasive developmental disorder not otherwise specified. *PLoS One* **8**, e76993.
- De Palma G, Blennerhassett P, Lu J, Park AJ, Philip V, Silva MA, Verdu EF, Collins SM & Bercik P (2012). Su1990 The role of microbiota in the maternal separation model of depression. *Gastroenterology* **142**(5S1), pS554.
- De Palma G, Nadal I, Medina M, Donat E, Ribes-Koninckx C, Calabuig M & Sanz Y (2010). Intestinal dysbiosis and reduced immunoglobulin-coated bacteria associated with coeliac disease in children. *BMC Microbiol* **10**, 63.
- Desbonnet L, Garrett L, Clarke G, Kiely B, Cryan JF & Dinan TG (2010). Effects of the probiotic *Bifidobacterium infantis* in the maternal separation model of depression. *Neuroscience* **170**, 1179–1188.
- Diaz–Heijtjz R, Wang S, Anuar F, Qian Y, Björkholm B, Samuelsson A, Hibberd ML, Forssberg H & Pettersson S (2011). Normal gut microbiota modulates brain development and behavior. *Proc Natl Acad Sci U S A* **108**, 3047–3052.
- Distrutti E, Cipriani S, Mencarelli A, Renga B & Fiorucci S (2013). Probiotics VSL#3 protect against development of visceral pain in murine model of irritable bowel syndrome. *PLoS One* **8**, e63893.
- Eutamene H & Bueno L (2007). Role of probiotics in correcting abnormalities of colonic flora induced by stress. *Gut* **56**, 1495–1497.
- Eutamene H, Lamine F, Chabo C, Theodorou V, Rochat F, Bergonzelli GE, Corthésy-Theulaz I, Fioramonti J & Bueno L (2007). Synergy between *Lactobacillus paracasei* and its bacterial products to counteract stress-induced gut permeability and sensitivity increase in rats. *J Nutr* **137**, 1901–1907.
- Felice VD, Gibney SM, Gosselin RD, Dinan TG, O’Mahony SM & Cryan JF (2014). Differential activation of the prefrontal cortex following psychological stress and colorectal distension in the maternally separated rat. *Neuroscience* **267**, 252–262.
- Finegold SM, Dowd SE, Gontcharova V, Liu C, Henley KE, Wolcott RD, Youn E, Summanen PH, Granpeesheh D, Dixon D, Liu M, Molitoris DR & Green JA 3rd (2010). Pyrosequencing study of fecal microflora of autistic and control children. *Anaerobe* **16**, 444–453.
- Finegold SM, Downes J & Summanen PH (2012). Microbiology of regressive autism. *Anaerobe* **18**, 260–262.
- García-Ródenas CL, Bergonzelli GE, Nutten S, Schumann A, Cherbut C, Turini M, Ornstein K, Rochat F & Corthésy-Theulaz I (2006). Nutritional approach to restore impaired intestinal barrier function and growth after neonatal stress in rats. *J Pediatr Gastroenterol Nutr* **43**, 16–24.
- Gareau MG, Jury J, MacQueen G, Sherman PM & Perdue MH (2007a). Probiotic treatment of rat pups normalises corticosterone release and ameliorates colonic dysfunction induced by maternal separation. *Gut* **56**, 1522–1528.
- Gareau MG, Jury J & Perdue MH (2007b). Neonatal maternal separation of rat pups results in abnormal cholinergic regulation of epithelial permeability. *Am J Physiol Gastrointest Liver Physiol* **293**, G198–G203.
- Gareau MG, Jury J, Yang PC, MacQueen G & Perdue MH (2006). Neonatal maternal separation causes colonic dysfunction in rat pups including impaired host resistance. *Pediatr Res* **59**, 83–88.
- Gareau MG, Silva MA & Perdue MH (2008). Pathophysiological mechanisms of stress-induced intestinal damage. *Curr Mol Med* **8**, 274–281.
- Geuking MB, Cahenzli J, Lawson MA, Ng DC, Slack E, Hapfelmeier S, McCoy KD & Macpherson AJ (2011). Intestinal bacterial colonization induces mutualistic regulatory T cell responses. *Immunity* **34**, 794–806.
- Gustafsson BE, Midtvedt T & Strandberg K (1970). Effects of microbial contamination on the cecum enlargement of germfree rats. *Scand J Gastroenterol* **5**, 309–314.
- Hansen CH, Nielsen DS, Kverka M, Zakostelska Z, Klimesova K, Hudcovic T, Tlaskalova-Hogenova H & Hansen AK (2012). Patterns of early gut colonization shape future immune responses of the host. *PLoS One* **7**, e34043.
- Hapfelmeier S, Lawson MA, Slack E, Kirundi JK, Stoel M, Heikenwalder M, Cahenzli J, Velykoredko Y, Balmer ML, Endt K, Geuking MB, Curtiss R 3rd, McCoy KD & Macpherson AJ (2010). Reversible microbial colonization of germ-free mice reveals the dynamics of IgA immune responses. *Science* **328**, 1705–1709.
- Heitkemper MM, Cain KC, Burr RL, Jun SE & Jarrett ME (2011). Is childhood abuse or neglect associated with symptom reports and physiological measures in women with irritable bowel syndrome? *Biol Res Nurs* **13**, 399–408.
- Hsiao EY, McBride SW, Hsien S, Sharon G, Hyde ER, McCue T, Codelli JA, Chow J, Reisman SE, Petrosino JF, Patterson PH & Mazmanian SK (2013). Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. *Cell* **155**, 1451–1463.
- Jeffery IB, Claesson MJ, O’Toole PW & Shanahan F (2012). Categorization of the gut microbiota: enterotypes or gradients? *Nat Rev Microbiol* **10**, 591–592.
- Kang DW, Park JG, Ilhan ZE, Wallstrom G, Labaer J, Adams JB & Krajmalnik-Brown R (2013). Reduced incidence of *Prevotella* and other fermenters in intestinal microflora of autistic children. *PLoS One* **8**, e68322.
- Konturek PC, Brzozowski T & Konturek SJ (2011). Stress and the gut: pathophysiology, clinical consequences, diagnostic approach and treatment options. *J Physiol Pharmacol* **62**, 591–599.
- Kunii J, Takahashi K, Kasakura K, Tsuda M, Nakano K, Hosono A & Kaminogawa S (2011). Commensal bacteria promote migration of mast cells into the intestine. *Immunobiology* **216**, 692–697.
- Ladd CO, Huot RL, Thirivikraman KV, Nemeroff CB, Meaney MJ & Plotsky PM (2000). Long-term behavioral and neuroendocrine adaptations to adverse early experience. *Prog Brain Res* **122**, 81–103.
- Lagier JC, Million M, Hugon P, Armougom F & Raoult D (2012). Human gut microbiota: repertoire and variations. *Front Cell Infect Microbiol* **2**, 136.
- Larsson MB, Tillisch K, Craig AD, Engstrom M, Labus J, Naliboff B, Lundberg P, Ström M, Mayer EA & Walter SA (2012). Brain responses to visceral stimuli reflect visceral sensitivity thresholds in patients with irritable bowel syndrome. *Gastroenterology* **142**, 463–472.

- Ledochowski M, Sperner-Unterweger B & Fuchs D (1998a). Lactose malabsorption is associated with early signs of mental depression in females: a preliminary report. *Dig Dis Sci* **43**, 2513–2517.
- Ledochowski M, Sperner-Unterweger B, Widner B & Fuchs D (1998b). Fructose malabsorption is associated with early signs of mental depression. *Eur J Med Res* **3**, 295–298.
- Ledochowski M, Widner B, Murr C, Sperner-Unterweger B & Fuchs D (2001). Fructose malabsorption is associated with decreased plasma tryptophan. *Scand J Gastroenterol* **36**, 367–371.
- Ledochowski M, Widner B, Sperner-Unterweger B, Propst T, Vogel W & Fuchs D (2000). Carbohydrate malabsorption syndromes and early signs of mental depression in females. *Dig Dis Sci* **45**, 1255–1259.
- Ley RE, Hamady M, Lozupone C, Turnbaugh PJ, Ramey RR, Bircher JS, Schlegel ML, Tucker TA, Schrenzel MD, Knight R & Gordon JI (2008). Evolution of mammals and their gut microbes. *Science* **320**, 1647–1651.
- Li M, Xue X, Shao S, Shao F & Wang W (2013). Cognitive, emotional and neurochemical effects of repeated maternal separation in adolescent rats. *Brain Res* **1518**, 82–90.
- Lippmann M, Bress A, Nemeroff CB, Plotsky PM & Monteggia LM (2007). Long-term behavioural and molecular alterations associated with maternal separation in rats. *Eur J Neurosci* **25**, 3091–3098.
- Lucassen PJ, Pruessner J, Sousa N, Almeida OF, Van Dam AM, Rajkowska G, Swaab DF & Czeh B (2014). Neuropathology of stress. *Acta Neuropathol* **127**, 109–135.
- Lyte M (2011). Probiotics function mechanistically as delivery vehicles for neuroactive compounds: microbial endocrinology in the design and use of probiotics. *Bioessays* **33**, 574–581.
- Lyte M, Vulchanova L & Brown DR (2011). Stress at the intestinal surface: catecholamines and mucosa–bacteria interactions. *Cell Tissue Res* **343**, 23–32.
- Macpherson AJ, Geuking MB & McCoy KD (2012). Homeland security: IgA immunity at the frontiers of the body. *Trends Immunol* **33**, 160–167.
- Matsumoto M, Kibe R, Ooga T, Aiba Y, Sawaki E, Koga Y & Benno Y (2013). Cerebral low-molecular metabolites influenced by intestinal microbiota: a pilot study. *Front Syst Neurosci* **7**, 9.
- Mayer EA (2011). Gut feelings: the emerging biology of gut–brain communication. *Nat Rev Neurosci* **12**, 453–466.
- Mayer EA, Savidge T & Shulman RJ (2014). Brain–gut microbiome interactions and functional bowel disorders. *Gastroenterology* **146**, 1500–1512.
- Mehdi S (2010). Antibiotic-induced psychosis: a link to D-alanine?. *Med Hypotheses* **75**, 676–677.
- Minot S, Bryson A, Chehoud C, Wu GD, Lewis JD & Bushman FD (2013). Rapid evolution of the human gut virome. *Proc Natl Acad Sci U S A* **110**, 12450–12455.
- Minot S, Sinha R, Chen J, Li H, Keilbaugh SA, Wu GD, Lewis JD & Bushman FD (2011). The human gut virome: inter-individual variation and dynamic response to diet. *Genome Res* **21**, 1616–1625.
- Moloney RD, O’Leary OF, Felice D, Bettler B, Dinan TG & Cryan JF (2012). Early-life stress induces visceral hypersensitivity in mice. *Neurosci Lett* **512**, 99–102.
- Mulle JG, Sharp WG & Cubells JF (2013). The gut microbiome: a new frontier in autism research. *Curr Psychiatry Rep* **15**, 337.
- Nadal I, Donat E, Ribes-Koninckx C, Calabuig M & Sanz Y (2007). Imbalance in the composition of the duodenal microbiota of children with coeliac disease. *J Med Microbiol* **56**, 1669–1674.
- Neufeld KM, Kang N, Bienenstock J & Foster JA (2011). Reduced anxiety-like behavior and central neurochemical change in germ-free mice. *Neurogastroenterol Motil* **23**, 255–264, e119.
- Nicholson JK, Holmes E, Kinross J, Burcelin R, Gibson G, Jia W & Pettersson S (2012). Host–gut microbiota metabolic interactions. *Science* **336**, 1262–1267.
- Ochoa-Repáraz J, Mielcarz DW, Begum-Haque S & Kasper LH (2011). Gut, bugs, and brain: role of commensal bacteria in the control of central nervous system disease. *Ann Neurol* **69**, 240–247.
- Oines E, Murison R, Mrdalj J, Grønli J & Milde AM (2012). Neonatal maternal separation in male rats increases intestinal permeability and affects behavior after chronic social stress. *Physiol Behav* **105**, 1058–1066.
- Olsson C & Holmgren S (2011). Autonomic control of gut motility: a comparative view. *Auton Neurosci* **165**, 80–101.
- Olszak T, An D, Zeissig S, Vera MP, Richter J, Franke A, Glickman JN, Siebert R, Baron RM, Kasper DL & Blumberg RS (2012). Microbial exposure during early life has persistent effects on natural killer T cell function. *Science* **336**, 489–493.
- O’Mahony SM, Hyland NP, Dinan TG & Cryan JF (2011). Maternal separation as a model of brain–gut axis dysfunction. *Psychopharmacology (Berl)* **214**, 71–88.
- O’Mahony SM, Marchesi JR, Scully P, Codling C, Ceolho AM, Quigley EM, Cryan JF & Dinan TG (2009). Early life stress alters behavior, immunity, and microbiota in rats: implications for irritable bowel syndrome and psychiatric illnesses. *Biol Psychiatry* **65**, 263–267.
- O’Malley D, Dinan TG & Cryan JF (2011). Altered expression and secretion of colonic interleukin-6 in a stress-sensitive animal model of brain–gut axis dysfunction. *J Neuroimmunol* **235**, 48–55.
- O’Malley D, Julio-Pieper M, Gibney SM, Dinan TG & Cryan JF (2010). Distinct alterations in colonic morphology and physiology in two rat models of enhanced stress-induced anxiety and depression-like behaviour. *Stress* **13**, 114–122.
- Park AJ, Collins J, Blennerhassett PA, Ghia JE, Verdu EF, Bercik P & Collins SM (2013). Altered colonic function and microbiota profile in a mouse model of chronic depression. *Neurogastroenterol Motil* **25**, 733–e575.
- Powell ND, Sloan EK, Bailey MT, Arevalo JM, Miller GE, Chen E, Kobor MS, Reader BF, Sheridan JF & Cole SW (2013). Social stress up-regulates inflammatory gene expression in the leukocyte transcriptome via β -adrenergic induction of myelopoiesis. *Proc Natl Acad Sci U S A* **110**, 16574–16579.
- Rabot S, Membrez M, Bruneau A, Gérard P, Harach T, Moser M, Raymond F, Mansourian R & Chou CJ (2010). Germ-free C57BL/6J mice are resistant to high-fat-diet-induced insulin resistance and have altered cholesterol metabolism. *FASEB J* **24**, 4948–4959.

- Rajilić-Stojanović M, Heilig HG, Tims S, Zoetendal EG & de Vos WM (2013). Long-term monitoring of the human intestinal microbiota composition. *Environ Microbiol* **15**, 1146–1159.
- Rhee SH, Pothoulakis C & Mayer EA (2009). Principles and clinical implications of the brain–gut–enteric microbiota axis. *Nat Rev Gastroenterol Hepatol* **6**, 306–314.
- Ringel Y & Maharshak N (2013). Intestinal microbiota and immune function in the pathogenesis of irritable bowel syndrome. *Am J Physiol Gastrointest Liver Physiol* **305**, G529–G541.
- Sandler RH, Finegold SM, Bolte ER, Buchanan CP, Maxwell AP, Väisänen ML, Nelson MN & Wexler HM (2000). Short-term benefit from oral vancomycin treatment of regressive-onset autism. *J Child Neurol* **15**, 429–435.
- Sapolsky RM, Romero LM & Munck AU (2000). How do glucocorticoids influence stress responses? Integrating permissive, suppressive, stimulatory, and preparative actions. *Endocr Rev* **21**, 55–89.
- Sekirov I, Russell SL, Antunes LC & Finlay BB (2010). Gut microbiota in health and disease. *Physiol Rev* **90**, 859–904.
- Slack E, Hapfelmeier S, Stecher B, Velykoredko Y, Stoel M, Lawson MA, Geuking MB, Beutler B, Tedder TF, Hardt WD, Bercik P, Verdu EF, McCoy KD & Macpherson AJ (2009). Innate and adaptive immunity cooperate flexibly to maintain host-microbiota mutualism. *Science* **325**, 617–620.
- Smith SM & Vale WW (2006). The role of the hypothalamic-pituitary-adrenal axis in neuroendocrine responses to stress. *Dialogues Clin Neurosci* **8**, 383–395.
- Söderholm JD & Perdue MH (2001). Stress and gastrointestinal tract. II. Stress and intestinal barrier function. *Am J Physiol Gastrointest Liver Physiol* **280**, G7–G13.
- Söderholm JD, Yates DA, Gareau MG, Yang PC, Macqueen G & Perdue MH (2002). Neonatal maternal separation predisposes adult rats to colonic barrier dysfunction in response to mild stress. *Am J Physiol Gastrointest Liver Physiol* **283**, G1257–G1263.
- Sommer F & Bäckhed F (2013). The gut microbiota – masters of host development and physiology. *Nat Rev Microbiol* **11**, 227–238.
- Sternbach H & State R (1997). Antibiotics: neuropsychiatric effects and psychotropic interactions. *Harv Rev Psychiatry* **5**, 214–226.
- Tannock GW & Savage DC (1974). Influences of dietary and environmental stress on microbial populations in the murine gastrointestinal tract. *Infect Immun* **9**, 591–598.
- Tannock GW & Smith JMB (1972). The effect of food and water deprivation (stress) on Salmonella-carrier mice. *J Med Microbiol* **5**, 283–289.
- Tillisch K & Labus JS (2011). Advances in imaging the brain–gut axis: functional gastrointestinal disorders. *Gastroenterology* **140**, 407–411.
- Tillisch K, Mayer EA & Labus JS (2011). Quantitative meta-analysis identifies brain regions activated during rectal distension in irritable bowel syndrome. *Gastroenterology* **140**, 91–100.
- Track NS (1980). The gastrointestinal endocrine system. *Can Med Assoc J* **122**, 287–292.
- van Tilburg MA, Palsson OS & Whitehead WE (2013). Which psychological factors exacerbate irritable bowel syndrome? Development of a comprehensive model. *J Psychosom Res* **74**, 486–492.
- Varghese AK, Verdu EF, Bercik P, Khan WI, Blennerhassett PA, Szechtman H & Collins SM (2006). Antidepressants attenuate increased susceptibility to colitis in a murine model of depression. *Gastroenterology* **130**, 1743–1753.
- Victor DW 3rd & Quigley EMM (2014). Hepatic encephalopathy involves interactions among the microbiota, gut, brain. *Clin Gastroenterol Hepatol* doi: 10.1016/j.cgh.2014.01.022.
- Wang L, Christophersen CT, Sorich MJ, Gerber JP, Angley MT & Conlon MA (2011). Low relative abundances of the mucolytic bacterium *Akkermansia muciniphila* and *Bifidobacterium* spp. in feces of children with autism. *Appl Environ Microbiol* **77**, 6718–6721.
- Wang L, Christophersen CT, Sorich MJ, Gerber JP, Angley MT & Conlon MA (2012). Elevated fecal short chain fatty acid and ammonia concentrations in children with autism spectrum disorder. *Dig Dis Sci* **57**, 2096–2102.
- Wang L, Christophersen CT, Sorich MJ, Gerber JP, Angley MT & Conlon MA (2013). Increased abundance of *Sutterella* spp. and *Ruminococcus torques* in feces of children with autism spectrum disorder. *Mol Autism* **4**, 42.
- Williams BL, Hornig M, Buie T, Bauman ML, Cho PM, Wick I, Bennett A, Jabado O, Hirschberg DL & Lipkin WI (2011). Impaired carbohydrate digestion and transport and mucosal dysbiosis in the intestines of children with autism and gastrointestinal disturbances. *PLoSOne* **6**, e24585.
- Williams BL, Hornig M, Parekh T & Lipkin WI (2012). Application of novel PCR-based methods for detection, quantitation, and phylogenetic characterization of *Sutterella* species in intestinal biopsy samples from children with autism and gastrointestinal disturbances. *MBio* **3**(1), e00261-11.
- Wostmann BS (1981). The germfree animal in nutritional studies. *Annu Rev Nutr* **1**, 257–279.
- Wu JC (2012). Psychological co-morbidity in functional gastrointestinal disorders: epidemiology, mechanisms and management. *J Neurogastroenterol Motil* **18**, 13–18.
- Yap IK, Angley M, Veselkov KA, Holmes E, Lindon JC & Nicholson JK (2010). Urinary metabolic phenotyping differentiates children with autism from their unaffected siblings and age-matched controls. *J Proteome Res* **9**, 2996–3004.
- Zoetendal EG, Rajilic-Stojanovic M & de Vos WM (2008). High-throughput diversity and functionality analysis of the gastrointestinal tract microbiota. *Gut* **57**, 1605–1615.

Additional information

Competing interests

None declared.