

Effects of Ginger Constituents on the Gastrointestinal Tract: Role of Cholinergic M₃ and Serotonergic 5-HT₃ and 5-HT₄ Receptors

Authors

Heinz H. Pertz¹, Jochen Lehmann^{2,3}, René Roth-Ehrang², Sigurd Elz⁴

Affiliations

¹ Institut für Pharmazie, Freie Universität Berlin, Berlin (Dahlem), Germany
² Institut für Pharmazie, Friedrich-Schiller-Universität Jena, Jena, Germany
³ College of Pharmacy, King Saud University, Riyadh, Saudi Arabia
⁴ Institut für Pharmazie, Universität Regensburg, Regensburg, Germany

Key words

- ginger
- *Zingiber officinale* Roscoe
- Zingiberaceae
- gastrointestinal diseases
- cholinergic M₃ receptors
- 5-HT₃ receptors
- 5-HT₄ receptors

Abstract

▼
 The herbal drug ginger (*Zingiber officinale* Roscoe) may be effective for treating nausea, vomiting, and gastric hypomotility. In these conditions, cholinergic M₃ receptors and serotonergic 5-HT₃ and 5-HT₄ receptors are involved. The major chemical constituents of ginger are [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol. We studied the interaction of [6]-gingerol, [8]-gingerol, [10]-gingerol (racemates), and [6]-shogaol with guinea pig M₃ receptors, guinea pig 5-HT₃ receptors, and rat 5-HT₄ receptors. In whole segments of guinea pig ileum (bioassay for contractile M₃ receptors), [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol slightly but significantly depressed the maximal carbachol response at an antagonist concentration of 10 μM. In the guinea pig myenteric plexus preparation (bioassay for contractile 5-HT₃ receptors), 5-HT maximal responses were depressed by [10]-gingerol from 93 ± 3% to 65 ± 6% at an antagonist concentration of 3 μM and to 48 ± 3% at an antagonist concentration of 5 μM following desensitization of 5-HT₄ receptors and blockade of 5-HT₁ and 5-HT₂ receptors. [6]-Shogaol (3 μM) induced depression to 61 ± 3%. In rat esophageal tunica muscularis mucosae (bioassay for relaxant 5-HT₄ receptors), [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol (2–6.3 μM) showed no agonist effects. The maximal 5-HT response remained unaffected in the presence of the compounds. It is concluded that the efficiency of ginger in reducing nausea and vomiting may be based on a weak inhibitory effect of gingerols and shogaols at M₃ and 5-HT₃ receptors. 5-HT₄ receptors, which play a role in gastroduodenal motility, appear not to be involved in the action of these compounds.

Introduction

▼
 In traditional medicine the powdered rhizome of ginger (*Zingiber officinale* Roscoe, Zingiberaceae) has been used for a long time to alleviate the symptoms of gastrointestinal diseases [1–3]. Clinical studies have shown that ginger is effective in reducing motion sickness, postoperative nausea and vomiting, hyperemesis gravidarum, and gastric hypomotility [4–7]. It has been hypothesized that the pharmacological activity of ginger lies in its pungent principles (gingerols and shogaols) and volatile oils (sesquiterpenes and monoterpenes) [8]. The major chemical constituents of the nonvolatile fraction of ginger are the arylalkanes [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol [3].

Attempts were made to characterize the receptor(s) that might be responsible for the therapeutic benefit of ginger on gastrointestinal function. The mechanism of action of ginger is

thought to lie in the ability of its active components to affect serotonin (5-HT) and muscarinic receptors in the gastrointestinal tract [9–11]. It has been shown that ginger possesses antiemetic properties against chemotherapy-induced vomiting in laboratory animals [12–14] as well as in humans [15]. The antiemetic effect of ginger was attributed to an antagonist interaction with 5-HT₃ receptors [14]. Interestingly, a contribution of the volatile oil to the 5-HT₃ receptor-mediated antiemetic effect of ginger has recently been suggested [16]. However, ginger was less effective in reducing cisplatin-induced emesis than the selective 5-HT₃ receptor antagonist granisetron [14]. Furthermore, ginger failed to prevent apomorphine-induced emesis, ruling out an involvement of dopamine receptors [14]. In addition, it has been suggested that the antiemetic effect of ginger is associated with an effect on gastric emptying; ginger and its active constituents [6]-gingerol, [8]-gingerol, [10]-

received October 18, 2010
 revised January 10, 2011
 accepted January 14, 2011

Bibliography

DOI <http://dx.doi.org/10.1055/s-0030-1270747>
 Published online February 8, 2011
 Planta Med 2011; 77: 973–978
 © Georg Thieme Verlag KG
 Stuttgart · New York ·
 ISSN 0032-0943

Correspondence

Prof. Dr. Heinz H. Pertz
 Institut für Pharmazie
 Freie Universität Berlin,
 Königin-Luise-Straße 2 + 4
 14195 Berlin (Dahlem)
 Germany
 Phone: + 49 30 83 85 31 35
 Fax: + 49 30 83 85 55 76
 hpertz@zedat.fu-berlin.de

gingerol, and [6]-shogaol have been reported to enhance gastric emptying of a charcoal meal in mice [17]. This finding is in line with the ability of ginger extracts to reverse cisplatin-induced delay in gastric emptying in rats [18]. The effect of ginger on gastric emptying was similar to that caused by metoclopramide, a 5-HT₄ receptor agonist with antagonist properties at 5-HT₃ receptors, and by ondansetron, a selective 5-HT₃ receptor antagonist [17,18]. Important evidence for the participation of 5-HT₄ receptors in gastric emptying induced by gastrointestinal prokinetics has been provided [19,20]. These findings unequivocally rule out any relevant contribution of 5-HT₃ receptor blockade in the pharmacological action of gastroprokinetics such as metoclopramide [21].

The aim of the present study was to evaluate the effectiveness of [6]-, [8]-, and [10]-gingerol and [6]-shogaol at cholinergic M₃ receptors in whole segments of guinea pig ileum, at serotonergic 5-HT₃ receptors of the longitudinal muscle-myenteric plexus preparation of guinea pig ileum, and at 5-HT₄ receptors of rat esophageal tunica muscularis mucosae. Based on the pharmacological profile of [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol, an antiemetic effect of ginger may be caused by the weak inhibitory properties of these compounds at the M₃ and 5-HT₃ receptors. The prokinetic properties of ginger are not mediated by a specific interaction with 5-HT₄ receptors.

Materials and Methods



Chemicals

[6]-Gingerol, [8]-gingerol, and [10]-gingerol (racemates) were synthesized in-house according to previously described standard procedures [22]. [6]-Shogaol was obtained from [6]-gingerol by dehydration. The purity of the compounds was >95% (analyzed by HPLC). The following drugs were obtained as gifts: methysergide hydrogen maleate (purity >99%) and tropisetron hydrochloride (purity >99%) (Novartis). The following compounds were purchased: atropine sulfate (purity >97%) and cocaine hydrochloride (purity >98.5%) (Merck); carbachol (purity >98%), corticosterone (purity >92%), 5-methoxytryptamine (purity 97%), pargyline hydrochloride (purity >98%), and SDZ-20557 hydrochloride (purity >99%) (Sigma-Aldrich); and choline chloride (purity >99%) and 5-hydroxytryptamine creatinine sulfate (purity >99%) (Janssen).

Animals

Animal studies were approved by Landesamt für Gesundheit und Soziales Berlin, Germany (ZH 12, T0035/00). All experimental procedures carried out in the present studies followed the guidelines of the Animals (Scientific Procedures) Act 1986. Male Wistar rats (250–300 g) and guinea pigs (250–450 g) were housed in plastic cages in a special temperature-controlled room (22 ± 2 °C, 50% humidity) on a 12:12-h light/dark cycle (light on at 07:00). They were allowed *ad libitum* access to food and water. Guinea pigs were stunned by a blow to the neck and bled. Rats were killed by decapitation following asphyxiation with CO₂.

Cholinergic M₃ receptor assay (guinea pig ileum)

Whole segments of guinea pig ileum (1.5 cm in length) were dissected and suspended in a water-jacketed 20-mL organ bath that contained Tyrode solution of the following composition (mM): NaCl 137, KCl 2.7, CaCl₂ 1.8, MgCl₂ 1.0, NaH₂PO₄ 0.4, NaHCO₃ 11.0, and glucose 5.6. The solution was gassed with 95% O₂/5%

CO₂ and warmed to a constant temperature of 37 °C (pH 7.4). One end of the segment was fixed on the bottom of the organ bath and the other was attached to an isotonic lever for continuous measurement of length changes. Preload at the beginning of the experiment was 7.5 mN. After a stabilization period of 20 min, the segments were stimulated three times with carbachol (1 μM) for a period of 45 min until the magnitude of the contractile response became constant. Four cumulative concentration-response curves were constructed at intervals of 15–20 min: the first curve to carbachol and the second through fourth curves to carbachol in the presence of antagonist or vehicle. Antagonists were incubated for 10 min. Previous studies by this group showed that successive concentration-response curves to carbachol in the absence of antagonist (control curves) were highly reproducible in guinea pig ileum (no differences in pEC₅₀ or E_{max} values). Contractile responses were expressed as a percentage value of the maximal response to carbachol in the first curve.

Serotonergic 5-HT₃ receptor assay (guinea pig myenteric plexus)

Strips of longitudinal muscle with adhering myenteric plexus from guinea pig ileum (1.5 cm in length) were prepared as previously described [23]. The strips were placed for the measurement of isometric changes in tension in 20-mL organ baths filled with Tyrode solution that additionally contained choline (1 μM) for *de novo* synthesis of acetylcholine, 5-methoxytryptamine (10 μM) for desensitization of 5-HT₄ receptors, and methysergide (1 μM) for blockade of 5-HT₁ and 5-HT₂ receptors [24]. The solution was continuously gassed with 95% O₂/5% CO₂ and warmed to a constant temperature of 37 °C (pH 7.4). The resting tension was adjusted to 7.5 mN at the beginning of each experiment. After a stabilization period of 30 min, the tissues were stimulated three times with 5-HT (3 μM) for a period of 60 min. Two noncumulative concentration-response curves were constructed at an interval of 60 min on each tissue by adding increasing concentrations of 5-HT (1–30 μM) to the organ bath. Each concentration was left in contact with the tissue for 15–30 s followed by washout with Tyrode solution (100 mL). The next higher concentration of 5-HT was administered 15 min later. Antagonists or vehicle were added 30 min before the second concentration-response curve and were retained in the bath fluid during construction of the second curve. Contractile responses were expressed as a percentage value of the maximal response to 5-HT in the first curve. Two noncumulative concentration-response curves to 5-HT on the same tissue (control curves) were highly reproducible, with the pEC₅₀ of the first curve (5.57 ± 0.02) being practically identical to that of the second curve (5.50 ± 0.06; *n* = 24). The E_{max} of the second 5-HT curve was 97 ± 1% relative to the E_{max} of the first curve.

Serotonergic 5-HT₄ receptor assay (rat esophageal tunica muscularis mucosae)

The entire rat esophagus was excised and the outer striated muscle layer of the tunica propria was removed by microdissection, leaving behind the inner smooth muscle tube of the tunica muscularis mucosae [25]. The muscle tube was divided into a proximal (cervical), middle (thoracic), and distal (abdominal) segment. From each segment, 2–3 strips were cut longitudinally and placed in Tyrode solution (pH 7.4, 37 °C) for measurement of isometric changes in tension. The Tyrode solution additionally contained methysergide (1 μM) to inhibit 5-HT₁ and 5-HT₂ receptors and cocaine (30 μM) and corticosterone (30 μM) to inhibit

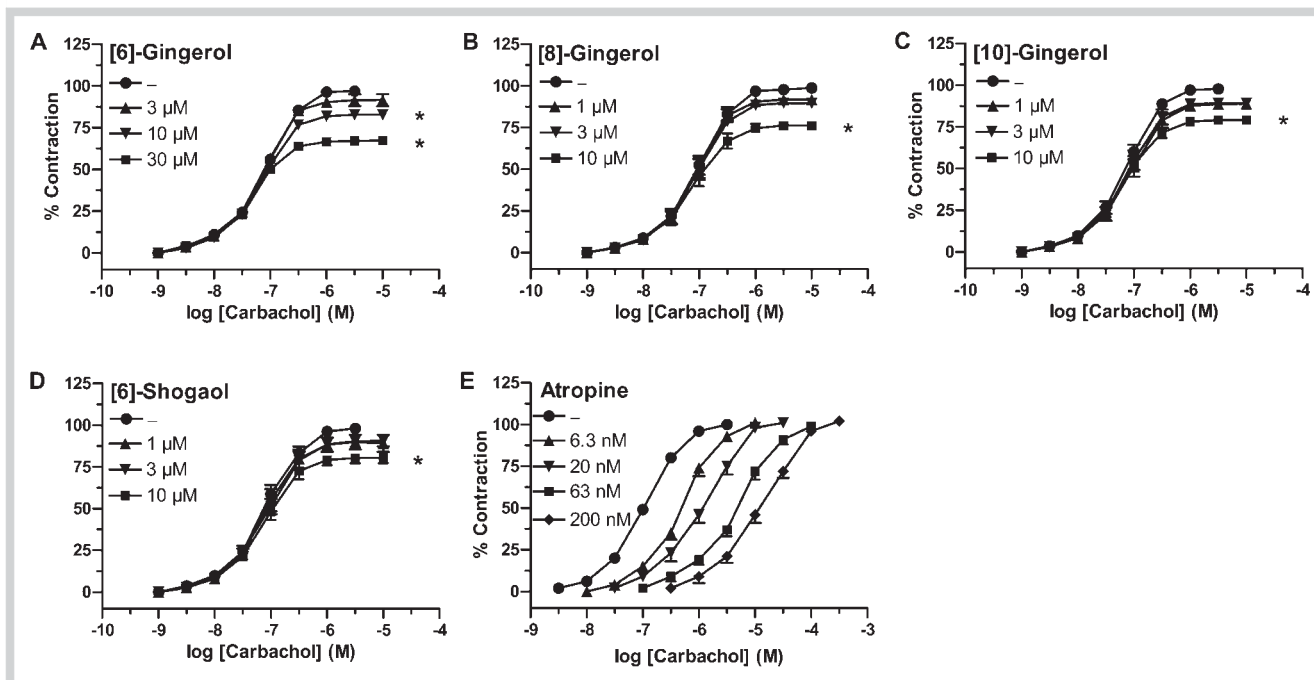


Fig. 1 Inhibition of carbachol-induced contraction by [6]-gingerol (A), [8]-gingerol (B), [10]-gingerol (C), [6]-shogaol (D), and atropine (M_3 receptor antagonist; E) in guinea pig ileum. Shown are cumulative concentration-re-

sponse curves to carbachol in the absence and presence of antagonist. Data are the mean \pm SEM from 4–6 animals; * $p < 0.05$.

neuronal and cellular uptake of 5-HT. The resting tension was adjusted to 1.5 mN at the beginning of each experiment. During a stabilization period of 30 min, the tissues were exposed to pargyline (100 μ M) to inhibit monoamine oxidases. The tissues were allowed to equilibrate for a further 30 min. After the stabilization and equilibration period, during which the tissues were washed with Tyrode solution (200 mL) every 15 min, the tissues were contracted with a submaximal concentration of carbachol (3 μ M) until the magnitude of the contractile response became constant (usually after 40 min). The relaxant response to 5-HT or arylalkanes was studied by constructing a cumulative concentration-response curve. For determination of antagonist affinity, a second concentration-response curve to 5-HT was performed following a further carbachol-induced (3 μ M) contraction. Antagonists or vehicle were incubated for 60 min. Effects were expressed as a percentage value of the maximal relaxation evoked by 5-HT in the first curve. Two cumulative concentration-response curves to 5-HT on the same tissue (control curves) were highly reproducible, with the pEC_{50} of the first curve (8.02 ± 0.07) being practically identical to that of the second curve (8.00 ± 0.06 ; $n = 15$). The E_{max} of the second 5-HT curve was $100 \pm 2\%$ relative to the E_{max} of the first curve.

Data presentation and statistical evaluation

Data are presented as the mean \pm SEM for n animals. Agonist potencies and maximal responses are expressed as pEC_{50} values (negative logarithm of the molar concentration of agonist producing 50% of the maximal response) and E_{max} values, respectively. In cases where an antagonist caused a parallel shift of an agonist curve, a Schild plot of $\log(\text{concentration ratio} - 1)$ versus $\log(\text{antagonist concentration})$ was constructed, and a pA_2 was calculated from the x -intercept of this plot. A slope of the Schild plot of 1 indicated competitive antagonism.

Where appropriate, differences between means were determined by Student's t -test (two-tailed), after checking the homogeneity of the variances. P values of less than 0.05 were considered to indicate a significant difference between the responses being compared.

Results

The effects of [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol on cholinergic M_3 receptors are shown in **Fig. 1 A–D**. In contrast to the effect of the reference antagonist atropine, which behaved as a competitive M_3 receptor antagonist ($pA_2 = 8.81 \pm 0.03$, slope of the Schild plot = 1.05 ± 0.03 ; **Fig. 1 E**), [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol (1–30 μ M) failed to evoke a rightward shift of the concentration-response curve to carbachol. However, the E_{max} values of carbachol were significantly depressed at an antagonist concentration of 10 μ M. Thus, ginger ingredients affected cholinergic M_3 receptors only at high concentrations and in a noncompetitive manner. [6]-Gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol did not evoke a contractile response via stimulation of cholinergic M_3 receptors. The effects of [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol on serotonergic 5-HT $_3$ receptors are shown in **Fig. 2 A–D**. The 5-HT $_3$ receptor-mediated contraction of guinea pig ileum by 5-HT is an indirect response that is based on the release of acetylcholine from intramural cholinergic nerves in the myenteric plexus followed by activation of cholinergic M_3 receptors on the longitudinal smooth muscle cell. At concentrations not interfering with M_3 receptors (1–6.3 μ M), there was no difference in the pEC_{50} values of 5-HT in the absence or presence of [6]-shogaol, [6]-gingerol, [8]-gingerol, and [10]-gingerol. However, E_{max} values were significantly depressed in the presence of

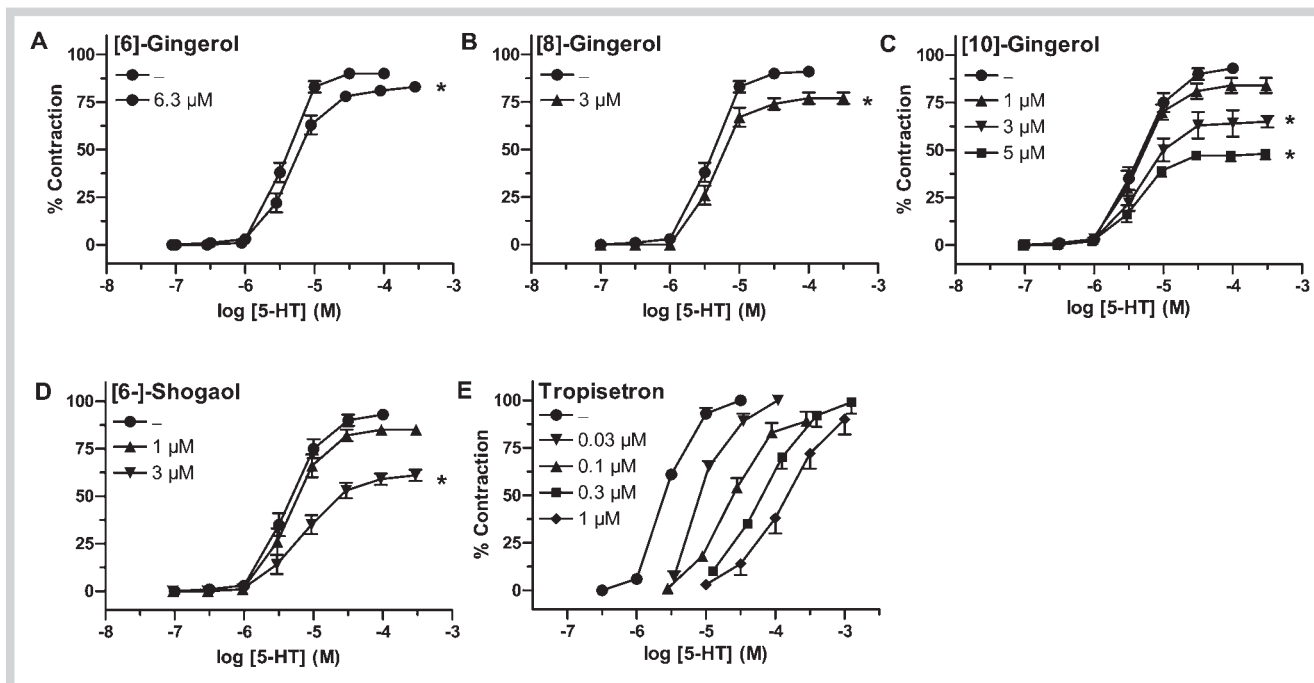


Fig. 2 Inhibition of 5-HT-induced contraction by [6]-gingerol (A), [8]-gingerol (B), [10]-gingerol (C), [6]-shogaol (D), and tropisetron (selective 5-HT₃ receptor antagonist; E) in the longitudinal muscle-myenteric plexus preparation of guinea pig ileum following desensitization of 5-HT₄ receptors and

blockade of 5-HT₁ and 5-HT₂ receptors (see Materials and Methods). Shown are noncumulative concentration-response curves to 5-HT (second curves) in the absence and presence of antagonist. Data are the mean \pm SEM from 4–6 animals. * $p < 0.05$.

the compounds. Most potent was [10]-gingerol, which concentration-dependently decreased the maximal 5-HT response from $93 \pm 3\%$ (control) to $65 \pm 6\%$ at an antagonist concentration of $3 \mu\text{M}$ and to $48 \pm 3\%$ at an antagonist concentration of $5 \mu\text{M}$. [6]-Shogaol ($3 \mu\text{M}$) showed similar antagonist effects by decreasing the maximal 5-HT response to $61 \pm 3\%$. Thus, gingerols and [6]-shogaol affected 5-HT₃ receptor-mediated ileal contractions in a noncompetitive manner. [6]-Gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol did not induce a contractile effect via stimulation of 5-HT₃ receptors. Tropicsetron, which was used as a reference antagonist, behaved as a competitive 5-HT₃ receptor antagonist ($pA_2 = 7.84 \pm 0.03$, slope of the Schild plot = 1.00 ± 0.04 ; **Fig. 2E**). The effects of [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol on serotonergic 5-HT₄ receptors are shown in **Fig. 3A–D**. The compounds failed to induce a relaxant response via stimulation of 5-HT₄ receptors of rat esophageal tunica muscularis mucosae. In the presence of the compounds (2 – $6.3 \mu\text{M}$), there was no rightward shift of the concentration-response curve to 5-HT. SDZ-205557, which was used as a reference antagonist, showed competitive 5-HT₄ receptor antagonist properties ($pA_2 = 7.34 \pm 0.05$, slope of the Schild plot = 0.95 ± 0.06 ; **Fig. 3E**).

Discussion

It is well established that 5-HT₃ receptor antagonists are efficient in the treatment of emesis induced by cancer chemotherapy and radiotherapy and in the prevention and treatment of postoperative nausea and vomiting [26]. From the present study, it is possible that the efficiency of the herbal drug ginger against chemotherapy-induced emesis [14, 15] and postoperative nausea and vomiting [5], at least at high dosage, may be due to an interaction

with 5-HT₃ receptors. In contrast to selective 5-HT₃ receptor antagonists such as tropisetron, which has been shown to competitively block 5-HT₃ receptor-mediated contractions in guinea pig ileum (**Fig. 2E**), [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol failed to produce a rightward shift of the concentration-response curve to 5-HT (**Fig. 2A–D**). A significant attenuation of the 5-HT maximal response in the presence of [8]-gingerol, [10]-gingerol, and [6]-shogaol was detected at an antagonist concentration of $3 \mu\text{M}$ (**Fig. 2A–D**). The noncompetitive inhibitory effects of the compounds may be mediated by binding to a modulatory site distinct from that of 5-HT. Our results are in accordance with recently published studies that demonstrated a weak inhibitory effect of ginger constituents via a noncompetitive blockade of 5-HT₃ receptors [11, 27]. However, it should be mentioned that arguments against the effectiveness of ginger in reducing the incidence of postoperative nausea and vomiting have also been presented [28]. Further studies are needed to substantiate the antiemetic properties of ginger via a blockade of 5-HT₃ receptors.

A current therapeutic application for ginger is motion sickness, although the effectiveness of the herbal drug for this indication seems to be controversial [4, 29]. In general, anticholinergic agents such as scopolamine are useful in the prevention and treatment of motion sickness. Due to the inability of ondansetron to be effective against motion sickness in humans, it has been concluded that 5-HT₃ receptors are not involved in the neural pathways that bring about motion sickness [30]. This finding suggests that any potential activity of ginger against motion sickness would be independent from the 5-HT₃ receptor blockade and might instead be based on a blockade of muscarinic receptors. Our study showed that ginger ingredients slightly but significantly affected cholinergic M₃ receptors only at high concentra-

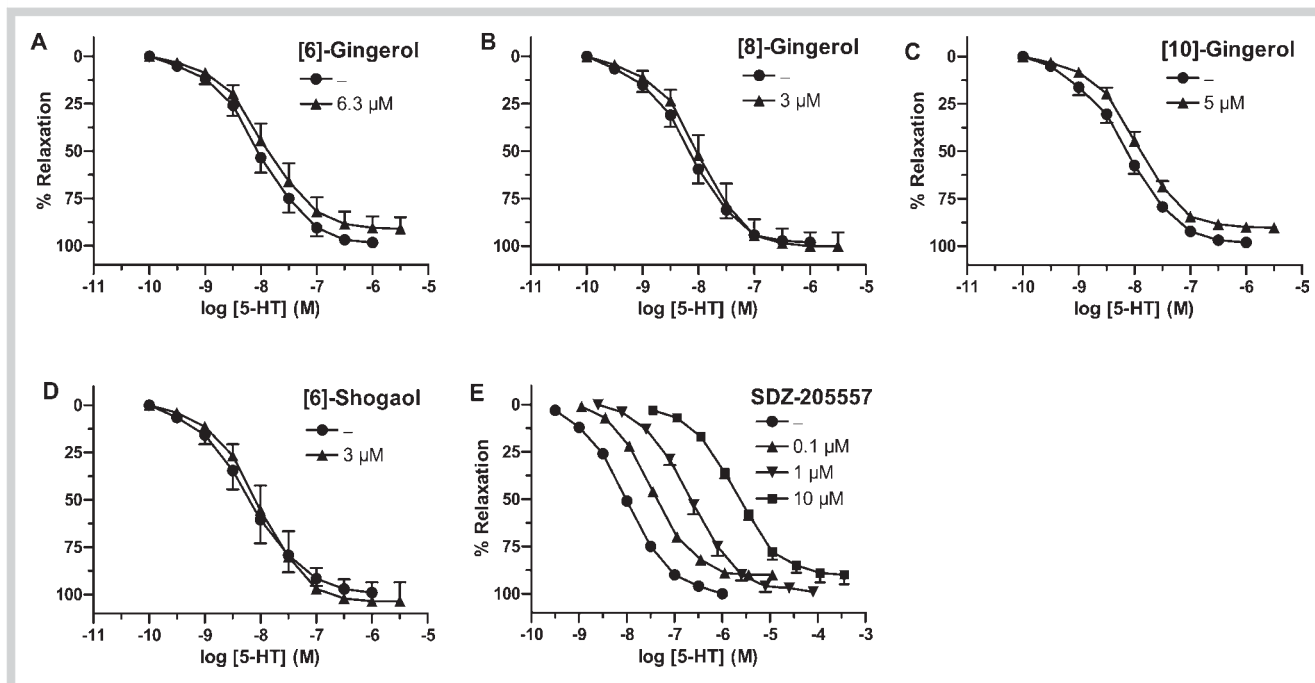


Fig. 3 Effects of [6]-gingerol (A), [8]-gingerol (B), [10]-gingerol (C), [6]-shogaol (D), and SDZ-205557 (selective 5-HT₄ receptor antagonist; E) on 5-HT-induced relaxation in rat esophageal tunica muscularis mucosae. Shown are

cumulative concentration-response curves to 5-HT (second curves) in the absence and presence of antagonist. Data are the mean \pm SEM from 4 animals.

tions. Our results are in accordance with the results of other studies [10, 11]. Surprisingly, it was recently shown that a ginger extract had an agonistic effect on cholinergic M₃ receptors [31]. This observation is in contrast to our results obtained with [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol.

A further aspect which is uncertain is the improvement of gastric emptying induced by ginger. Micklefield et al. [7] reported that oral ginger improved gastroduodenal motility, while Phillips et al. [32] found that ginger had no effect on gastric emptying. Efforts were made to elucidate the mode of action underlying the potential prokinetic effect of ginger. It was suggested that the enhancement of gastrointestinal motility by ginger is mediated by a blockade of 5-HT₃ receptors, since the effect was similar to that caused by metoclopramide and ondansetron [17, 18]. It should be emphasized, however, that the gastrointestinal prokinetic activity of metoclopramide is based on the agonist properties of this drug at 5-HT₄ receptors [33]. 5-HT₄ receptor activation facilitates acetylcholine release from cholinergic neurons, thereby triggering the peristaltic reflex [34]. The present study on ginger rules out any interaction of ginger constituents with 5-HT₄ receptors, which are the responsible targets for serotonergic prokinetic agents. Our study shows that [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol failed to possess agonist activity at 5-HT₄ receptors. Therefore, a role for ginger in stimulating gastrointestinal motility via 5-HT₄ receptors is unlikely. Interestingly, it was recently demonstrated that ginger failed to improve delayed gastrointestinal transit in rat postoperative ileus, a process in which the 5-HT₄ receptor is partly involved [35].

In conclusion, our data support the possibility that the herbal drug ginger may be beneficial in reducing nausea and vomiting due to the weak inhibition of cholinergic M₃ receptors and serotonergic 5-HT₃ receptors by gingerols and shogaols. However, a prokinetic effect of gingerols and shogaols via stimulation of

5-HT₄ receptors can be excluded. Admittedly, a prokinetic effect of ginger might be mediated by other pathways, such as the inhibition of L-type Ca²⁺ channels [36]. [6]-Gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol have in common that they inhibited agonist responses to carbachol and 5-HT in a noncompetitive manner (● Figs. 1A–D and 2A–D) in contrast to the reference antagonists (atropine and tropisetron), which showed competitive antagonism (○ Figs. 1E and 2E). The present study has certain limitations. Most notably, we tested the racemates of [6]-gingerol, [8]-gingerol, and [10]-gingerol instead of the naturally occurring enantiomer. It would be particularly interesting to clarify whether the stereochemistry of natural occurring gingerols plays an essential role in the antiemetic activity of ginger. This may not be the case, since the achiral component of ginger, [6]-shogaol, was at least equally potent to gingerols in inhibiting 5-HT₃ and M₃ receptor function in this and in other studies [9, 11, 27]. In addition, our studies were restricted to the pharmacological profile of [6]-gingerol, [8]-gingerol, [10]-gingerol, and [6]-shogaol. However, other nonvolatile constituents of ginger (e.g., diacetyl-[6]-gingerdiol and [6]-dehydrogingerdione) and volatile oil constituents have also been demonstrated to possess 5-HT₃ receptor-blocking properties [16, 27]. Nonetheless, the current data support the finding that ginger is useful against nausea and vomiting [37].

Acknowledgements

▼ The generous gifts of compounds by the pharmaceutical companies mentioned in Materials and Methods are gratefully acknowledged.

References

- 1 Afzal M, Al-Hadidi D, Menon M, Pesek J, Dhimi MS. Ginger: an ethnomedical, chemical and pharmacological review. *Drug Metabol Drug Interact* 2001; 18: 159–190
- 2 Chrubasik S, Pittler MH, Roufogalis BD. *Zingiberis* rhizoma: a comprehensive review on the ginger effect and efficacy profiles. *Phytomedicine* 2005; 12: 684–701
- 3 Ali BH, Blunden G, Tanira MO, Nemmar A. Some phytochemical, pharmacological and toxicological properties of ginger (*Zingiber officinale* Roscoe): a review of recent research. *Food Chem Toxicol* 2008; 46: 409–420
- 4 Mowrey DB, Clayson DE. Motion sickness, ginger, and psychosis. *Lancet* 1982; 1: 655–657
- 5 Phillips S, Ruggier R, Hutchinson SE. *Zingiber officinale* (ginger) – an antiemetic for day case surgery. *Anaesthesia* 1993; 48: 715–717
- 6 Fischer-Rasmussen W, Kjaer SK, Dahl C, Asing U. Ginger treatment of hyperemesis gravidarum. *Eur J Obstet Gynecol Reprod Biol* 1991; 38: 19–24
- 7 Micklefield GH, Redeker Y, Meister V, Jung O, Greving I, May B. Effects of ginger on gastroduodenal motility. *Int J Clin Pharmacol Ther* 1999; 37: 341–346
- 8 Kemper KJ. Ginger (*Zingiber officinale*). The Longwood Herbal Task Force, 1999. Available at <http://www.longwoodherbal.org/ginger/ginger.pdf>. Accessed January 31, 2011
- 9 Yamahara J, Rong HQ, Iwamoto M, Kobayashi G, Hisashi Matsuda H, Fujimura H. Active components of ginger exhibiting anti-serotonergic action. *Phytother Res* 1989; 3: 70–71
- 10 Lien HC, Sun WM, Chen YH, Kim H, Hasler W, Owyang C. Effects of ginger on motion sickness and gastric slow-wave dysrhythmias induced by circular vection. *Am J Physiol Gastrointest Liver Physiol* 2003; 284: G481–G489
- 11 Abdel-Aziz H, Windeck T, Ploch M, Verspohl EJ. Mode of action of gingerols and shogaols on 5-HT₃ receptors: binding studies, cation uptake by the receptor channel and contraction of isolated guinea-pig ileum. *Eur J Pharmacol* 2006; 530: 136–143
- 12 Yamahara J, Rong HQ, Naitoh Y, Kitani T, Fujimura H. Inhibition of cytotoxic drug-induced vomiting in suncus by a ginger constituent. *J Ethnopharmacol* 1989; 27: 353–355
- 13 Kawai T, Kinoshita K, Koyama K, Takahashi K. Anti-emetic principles of *Magnolia obovata* bark and *Zingiber officinale* rhizome. *Planta Med* 1994; 60: 17–20
- 14 Sharma SS, Kochupillai V, Gupta SK, Seth SD, Gupta YK. Antiemetic efficacy of ginger (*Zingiber officinale*) against cisplatin-induced emesis in dogs. *J Ethnopharmacol* 1997; 57: 93–96
- 15 Pillai AK, Sharma KK, Gupta YK, Bakhshi S. Anti-emetic effect of ginger powder versus placebo as an add-on therapy in children and young adults receiving high emetogenic chemotherapy. *Pediatr Blood Cancer* 2011; 56: 234–238
- 16 Riyazi A, Hensel A, Bauer K, Geissler N, Schaaf S, Verspohl EJ. The effect of the volatile oil from ginger rhizomes (*Zingiber officinale*), its fractions and isolated compounds on the 5-HT₃ receptor complex and the serotonergic system of the rat ileum. *Planta Med* 2007; 73: 355–362
- 17 Yamahara J, Huang Q, Li Y, Xu L, Fujimura H. Gastrointestinal motility enhancing effect of ginger and its active constituents. *Chem Pharm Bull* 1990; 38: 430–431
- 18 Sharma SS, Gupta YK. Reversal of cisplatin-induced delay in gastric emptying in rats by ginger (*Zingiber officinale*). *J Ethnopharmacol* 1998; 62: 49–55
- 19 Gale JD, Reeves JJ, Bunce KT. Antagonism of the gastroprokinetic effect of metoclopramide by GR 125487, a potent and selective 5-HT₄ receptor antagonist. *J Gastrointest Motil* 1993; 5: 192
- 20 Hedge SS, Wong AG, Perry MR, Ku P, Moy TM, Loeb M, Eglen RM. 5-HT₄ receptor mediated stimulation of gastric emptying in rats. *Naunyn Schmiedebergs Arch Pharmacol* 1995; 351: 589–595
- 21 Tonini M. Recent advances in the pharmacology of gastrointestinal prokinetics. *Pharmacol Res* 1996; 33: 217–226
- 22 Demiff P, Macleod I, Whiting DA. Syntheses of the (±)-[n]-gingerols (pungent principles of ginger) and related compounds through regio-selective aldol condensations: relative pungency assays. *J Chem Soc [Perkin I]* 1981; 82–87
- 23 Buchheit KH, Engel G, Mutschler E, Richardson B. Study of the contractile effect of 5-hydroxytryptamine (5-HT) in the isolated longitudinal muscle strip from guinea-pig ileum. *Naunyn Schmiedebergs Arch Pharmacol* 1985; 329: 36–41
- 24 Craig DA, Eglen RM, Walsh LKM, Perkins LA, Whiting RL, Clarke DE. 5-Methoxytryptamine and 2-methyl-5-hydroxytryptamine-induced desensitization as a discriminative tool for the 5-HT₃ and putative 5-HT₄ receptors in guinea pig ileum. *Naunyn Schmiedebergs Arch Pharmacol* 1990; 342: 9–16
- 25 Akbarali HI, Bieger D, Triggler CR. Tetrodotoxin-sensitive and -insensitive relaxations in the rat oesophageal tunica muscularis mucosae. *J Physiol (London)* 1986; 381: 49–63
- 26 Veyrat-Follet C, Farinotti R, Palmer JL. Physiology of chemotherapy-induced emesis and antiemetic therapy. Predictive models for evaluation of new compounds. *Drugs* 1997; 53: 206–234
- 27 Abdel-Aziz H, Nahrstedt A, Petereit F, Windeck T, Ploch M, Verspohl EJ. 5-HT₃ receptor blocking activity of arylalkanes isolated from the rhizome of *Zingiber officinale*. *Planta Med* 2005; 71: 609–616
- 28 Visalyaputra S, Petchpaisit N, Somcharoen K, Choavaratana R. The efficacy of ginger root in the prevention of postoperative nausea and vomiting after outpatient gynaecological laparoscopy. *Anaesthesia* 1998; 53: 506–510
- 29 Stewart JJ, Wood MJ, Wood CD, Mims ME. Effects of ginger on motion sickness susceptibility and gastric function. *Pharmacology* 1991; 42: 111–120
- 30 Stott JR, Barnes GR, Wright RJ, Ruddock CJ. The effect on motion sickness and oculomotor function of GR 38032F, a 5-HT₃-receptor antagonist with anti-emetic properties. *Br J Clin Pharmacol* 1989; 27: 147–157
- 31 Ghayur MN, Khan AH, Gilani AH. Ginger facilitates cholinergic activity possibly due to blockade of muscarinic autoreceptors in rat stomach fundus. *Pakistan J Pharm Sci* 2007; 20: 231–235
- 32 Phillips S, Hutchinson S, Ruggier R. *Zingiber officinale* does not affect gastric emptying rate. A randomised, placebo-controlled, crossover trial. *Anaesthesia* 1993; 48: 393–395
- 33 Hoyer D, Clarke DE, Fozard JR, Hartig PR, Martin GR, Mylecharane EJ, Saxena PR, Humphrey PPA. VII. International Union of Pharmacology classification of receptors for 5-hydroxytryptamine (serotonin). *Pharmacol Rev* 1994; 46: 157–203
- 34 Bockaert J, Fozard JR, Dumuis A, Clarke DE. The 5-HT₄ receptor: a place in the sun. *Trends Pharmacol Sci* 1992; 13: 141–145
- 35 Tokita Y, Yuzurihara M, Sakaguchi M, Satoh K, Kase Y. The pharmacological effects of Daikenchuto, a traditional herbal medicine, on delayed gastrointestinal transit in rat postoperative ileus. *J Pharmacol Sci* 2007; 104: 303–310
- 36 Ghayur MN, Gilani AH. Species differences in the prokinetic effects of ginger. *Int J Food Sci Nutr* 2006; 57: 65–73
- 37 Ernst E, Pittler MH. Efficacy of ginger for nausea and vomiting: a systematic review of randomized clinical trials. *Br J Anaesth* 2000; 84: 367–371